Preliminary communication

2,3-Diacetamido-2,3-dideoxy-L-guluronic acid: a new acidic amino sugar from *Pseudomonas aeruginosa* O:3a,d,e lipopolysaccharide

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The O-specific polysaccharides of *P. aeruginosa* O:3 (Lanyi classification) possess similar structures that are composed of trisaccharide repeating-units containing 2-acetamido-2-deoxy-D-fucose and uncommon, acidic, diamino sugars¹. Recently, two of these sugars were identified as 2,3-diacetamido-2,3-dideoxy-D-mannuronic acid and its 2-imidazoline derivative². We now report the identification of 2,3-diacetamido-2,3-dideoxy-L-guluronic acid as a constituent of *P. aeruginosa* O:3a,d,e (strain 170006) O-specific polysaccharide.

The acidic polysaccharide (M_{GalA} 0.5; paper electrophoresis; pyridine acetate buffer, pH 4.5) was obtained by mild, acid degradation (1% CH₃CO₂H, 100°, 2 h) of the lipopolysaccharide isolated from dry bacterial cells by the Westphal procedure³. Hydrolvsis (4M HCl, 100°, 4 h) of the polysaccharide followed by conventional sugar analysis resulted in identification of 2-amino-2-deoxy-D-fucose in 3% yield as the single monosaccharide. The ¹³C-n.m.r. spectrum of the polysaccharide contained signals for three anomeric carbons (100.0, 99.7, and 98.0 p.p.m.), five carbons carrying nitrogen (57.8, 51.3, 50.8, 49.7, and 45.0 p.p.m.), one C-methyl group of a 6-deoxyhexose (16.4 p.p.m.), four acetamido methyl groups in the region 23.0-23.7 p.p.m., and six carbonyl groups in the region 174–177 p.p.m., as well as signals at 19.8 and 167.2 p.p.m. belonging to C-methyl and C-2, respectively, of a 2-methyl-2-imidazoline derivative². All other signals for the 2,3-(1-acetyl-2-methyl-2-imidazolino-5,4)-2,3-dideoxy-\(\beta\)-mannuronic acid residue (100.0, 76.3, 71.7, 57.8, and 51.3 p.p.m.) were also present in the spectrum. Signals for hydroxymethyl groups were absent. Therefore, it is proposed that the trisaccharide repeating-unit of the polysaccharide comprises 2-acetamido-2-deoxy-D-fucose, a diacetamidodideoxyuronic acid, and a 2-imidazoline derivative of a uronic acid probably having the manno configuration.

Solvolysis⁴ of the polysaccharide with hydrogen fluoride $(25^{\circ}, 3 \text{ h})$ gave the acidic trisaccharide 1, M_{GalA} 0.5, which was isolated by gel filtration on Sephadex G-15 in almost quantitative yield. The ¹³C-n.m.r. spectrum of 1 showed that the solvolysis had cleaved selectively the *N*-acetylfucosaminidic linkages. Thus, 1 was the chemical repeating-unit of the polysaccharide. Treatment of 1, in sequence, with borohydride, periodate, borohydride, and 5% aqueous triethylamine $(60^{\circ}, 3 \text{ h}; \text{to cleave the imidazoline ring})$

gave the acidic oligosaccharide 2, $M_{\rm GalA}$ 0.9. Carboxyl-reduction⁵ of 2 gave the neutral oligosaccharide 3, which was further acetylated to give 4.

The coupling constants $(J_{1'2}, 1.5, J_{2'3}, 3.6, J_{3'4}, 10.0, \text{ and } J_{4'5}, 10.0 \text{ Hz})$ determined from the 360-MHz ¹H-n.m.r. spectrum of 4 proved H-3',4',5' to be axial and H-2' to be equatorial and, thus, the configuration of the terminal hexose residue to be manno. The only coupling constants that could be determined for the second hexose residue were $J_{1,2}$ 3.5 and $J_{4,5}$ 2.6 Hz, the latter indicating H-4 to be equatorial. Comparison of the ¹³C-n.m.r. spectra of 3 and the analogous oligosaccharide obtained from the P. aeruginosa O:3a,d O-specific polysaccharide² revealed the presence of the same aglycon (2-acetamido-2-deoxythreitol) and terminal residue (2,3-diacetamido-2,3-dide $oxy-\beta$ -mannopyranose) and allowed their signals to be assigned. The remaining six signals in the spectrum of 3 (99.4, 76.0, 68.2, 61.8, 50.1, and 45.6 p.p.m.) should be assigned, therefore, to the penultimate hexose residue, those at 99.4 and 61.8 p.p.m. belonging unambiguously to C-1 and C-6, respectively. Furthermore, the signal at 76.0 p.p.m. was assigned to C-4, according to its downfield shift in the spectrum of 2 (to 77.4 p.p.m.), which is characteristic for the conversion of a hexose into the corresponding uronic acid⁶. Thus, the carbons carrying acetamido groups were C-2 and C-3 (signals at 45.6 and 50.1 p.p.m.). The remaining signal at 68.2 p.p.m. was consequently assigned to C-5. The C-1-H-1 coupling constant (J 172.0 Hz, determined from the ¹³C-n.m.r. spectrum of the polysaccharide) indicated the unknown 2,3-diacetamido-2,3-dideoxyuronic acid to be α -linked, assuming its 4C_1 conformation 7,8 . Therefore, the 13 C-n.m.r. data for 3 were compared with those for all eight methyl 2,3-diacetamido-2,3-dideoxy-α-D-hexopyranosides*. The comparison revealed the similarity in the chemical shifts of the signals for C-2 and C-5 in the spectra of 3 (45.6 and 68.2 p.p.m.) and methyl 2,3-diacetamido-2,3-dideoxy- α -D-gulopyranoside (45.4 and 68.0 p.p.m., respectively), and differences from the values for the other hexosides (\geqslant 3.5 p.p.m. for C-2 and 1 p.p.m. for C-5). Thus, the configuration of the unknown sugar is *gulo*.

Attempts to isolate 2,3-diamino-2,3-dideoxygulose by acid hydrolysis of 3 were unsuccessful due to its instability, which was supported by model experiments with methyl 2,3-diacetamido-2,3-dideoxy- α -D-gulopyranoside. In order to determine the absolute configuration of the new sugar, the dependence of the glycosidation shifts of aglycon carbon signals in the ¹³C-n.m.r. spectra on the absolute and anomeric configuration of the glycon⁹ was used. The relatively low (by module) effect (-1.4 p.p.m.) on C-4 of the 2-acetamido-2-deoxy-D-fucosyl residue in the spectrum of the polysaccharide caused by substitution at position 3 by a 2,3-diacetamido-2,3-dideoxy- α -gulopyranosyluronic acid residue indicated the L configuration of the sugar substituent. This assignment was supported by the $[\alpha]_D$ value of -66.7° (water) for 3.

Thus, the O-specific polysaccharide of *P. aeruginosa* O:3a,d,e contains 2,3-diacetamido-2,3-dideoxy-L-guluronic acid, which has not been observed previously in Nature, and the structure of the chemical repeating-unit of the polysaccharide is the trisaccharide 1.

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^{*}The synthesis of these compounds will be described elsewhere.